



**SITA** | **13° CONGRESSO NAZIONALE**  
Società Italiana di Terapia Antinfettiva  
Antibatterica Antivirale Antifungina  
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# Diagnostica e terapia dell'aspergillosi invasiva nel paziente ematologico

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**OSPEDALE POLICLINICO SAN MARTINO**  
Sistema Sanitario Regione Liguria  
Istituto di Ricovero e Cura a Carattere Scientifico

# DISCLOSURES

Lecture or board meeting, grants to my institution  
None related to this presentation

- Allovir
- bioMerieux
- Gilead
- Janssen
- Moderna
- Mundipharma
- Pfizer

# ID consult on IFD in hematology



- Why it may feel like mission impossible?

1. Haematology specialists may be more knowledgeable than ID in IFD in haematology/HSCT
2. They care deeply about their patients ... and they do not accept that infection might lead to death
3. Clinical signs and symptoms might be very limited
4. Definite diagnosis is difficult due to limited feasibility and sensitivity of biopsy ...
5. Once you put your foot down but make a mistake – regaining the trust is not easy ....

# Specifics of IFD in haematology

## Prophylaxis

High rate (>6-8%) of IMI in certain patient groups but not in others, thus **anti-mould prophylaxis strongly recommended in certain groups**

Breakthrough IFD present, also due to persistent immune deficit

# Anti-fungal prophylaxis

**Table 1** Established risk groups for IFD and recommended antifungal prophylaxis coverage in adults

Risk level	Risk groups	Recommended prophylaxis†
High risk >10% incidence of IFD	Neutrophil <0.1 × 10 <sup>9</sup> /L for >3 weeks or <0.5 × 10 <sup>9</sup> /L for >5 weeks (e.g. allogeneic HSCT) Corticosteroids >1 mg/kg prednisolone equivalent and neutrophils <1 × 10 <sup>9</sup> /L for >1 week Corticosteroids >2 mg/kg prednisolone equivalent >2 weeks Unrelated, mismatched or cord blood allogeneic HSCT GVHD – extensive or severe AML – induction/reinduction ALL – induction/reinduction MDS	First line: Posaconazole Alternate: Voriconazole
Low risk Less than 5% incidence of IFD	Autologous HSCT for mucositis Allogeneic neutrophil Lym	
Very low risk Less than 5% incidence of IFD No mucositis		

**Table 3.** ECIL recommendations on primary antifungal prophylaxis in adult patients with AML and MDS undergoing intensive remission-induction chemotherapy<sup>a</sup>

Antifungal agent	Grading	Comments
Posaconazole oral solution 200 mg q8h or tablet 300 mg q24h following a loading dose of 300 mg q12h on day 1	A-I	Recommended if baseline incidence of mould infections is high. Given the increased absorption of the tablet, it is likely that the need for therapeutic drug monitoring will become restricted to specific populations (e.g. severe mucositis).
Fluconazole 400 mg q24h	B-I	Only recommended if the incidence of mould infections is low. Fluconazole may be part of an integrated care strategy together with a mould-directed diagnostic approach.
Itraconazole oral solution 2.5 mg/kg q12h	B-I	Recommended if baseline incidence of mould infections is high. May be limited by drug–drug interactions or patient tolerability. It is recommended to monitor serum drug concentrations.
Voriconazole 200 mg q12h	B-II	Recommended if baseline incidence of mould infections is high. It is recommended to monitor serum drug concentrations.
All echinocandins	C-II	Insufficient data on efficacy and tolerability.
Liposomal amphotericin B	C-II	Insufficient data on dose, frequency and duration, as well as on efficacy and tolerability.
Lipid-associated amphotericin B	C-II	Insufficient data on dose, frequency and duration, as well as on efficacy and tolerability.
Aerosolized liposomal amphotericin B (10 mg twice weekly)	B-I	Only when combined with fluconazole 400 mg q24h.
Amphotericin B deoxycholate	A-II against	
Aerosolized amphotericin B deoxycholate	A-I against	

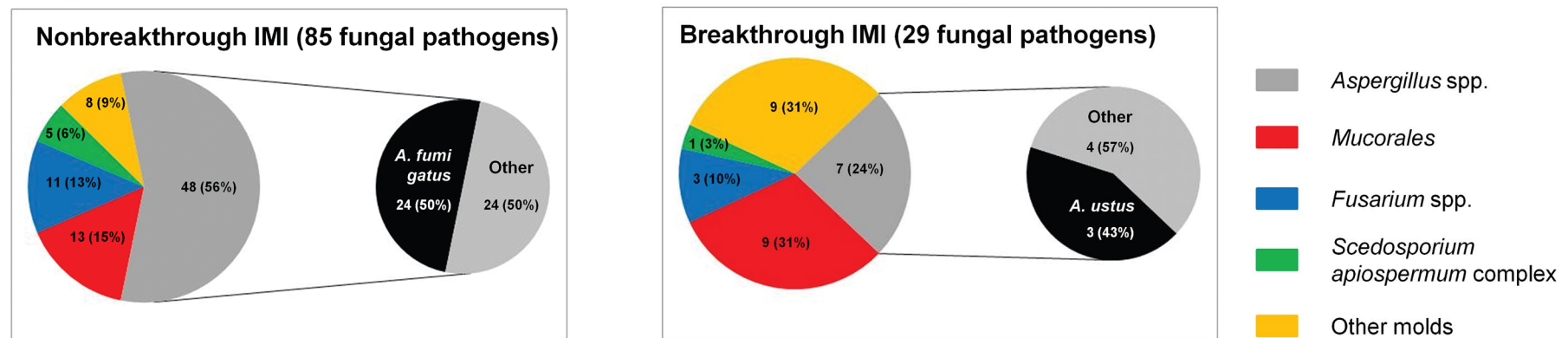
<sup>a</sup>Primary antifungal prophylaxis might be considered during intensified consolidation therapy (see text).

Treatment for solid org

# Breakthrough IFD

Usually approx. 3%, up to 12% in some cohorts

Pathogens: rare moulds and azole-resistant *Aspergillus* species



**Figure 1.** Distribution of fungal pathogens in breakthrough and nonbreakthrough invasive mold infections.

# Specifics of IFD in haematology

## Prophylaxis

High rate (>6-8%) of IMI in certain patient groups but not in others, thus anti-mould prophylaxis strongly recommended in certain groups  
Breakthrough IFD present, also due to persistent immune deficit

## Diagnosis

EORTC/MSG diagnostic criteria available for 20 years and updated, but not developed for daily clinical practice (e.g. bronchoinvasive aspergillosis – unclassified)

# 20 years of diagnosing invasive mould infections in the immunocompromised

- Risk factors + suggestive clinical presentation

EORTC/MSGERC criteria: 2002, 2008 and **2019** developed for **clinical research in the immunocompromised** at risk of IFD

- Probability levels of having invasive fungal infection:
  - **Proven** Histopathologic, cytopathologic, or direct microscopy in biopsy or culture from a sterile site
  - **Probable** Presence of Host factor + Clinical criterion + Mycology criterion
  - **Possible** Presence of Host factor + Clinical criterion

## Host factor

- Recent neutropenia
- HSCT incl. GvHD, **haematological malignancy, SOT**
- Steroids ( $\geq 0.3$  mg/kg for  $\geq 3$  weeks in the past 60 days)
- **Other T-cell immunosuppressants**
- B-cell immunosuppressants
- Inherited severe immunodeficiency



## Clinical criteria

- Dense, well-circumscribed lesion(s) with or without a halo sign
- Air crescent sign
- Cavity
- **Reverse halo sign**
- **Wedge-shape and segmental or lobar consolidations**



## Mycology criteria

- Direct microscopy, or **culture** in non-sterile samples (BAL, sputum, aspirate)
- **Indirect tests**
  - **GM**
    - Single serum GM  $\geq 1.0$
    - BAL GM  $\geq 1.0$
    - Single serum/plasma:  $\geq 0.7$  and BAL fluid  $\geq 0.8$
  - **Aspergillus PCR**
    - Plasma/serum/whole blood **2** or more consecutive PCR tests positive
    - BAL fluid 2 or more duplicate PCR tests positive
    - At least 1 PCR test positive in plasma/serum/whole blood and 1 PCR test positive in BAL fluid

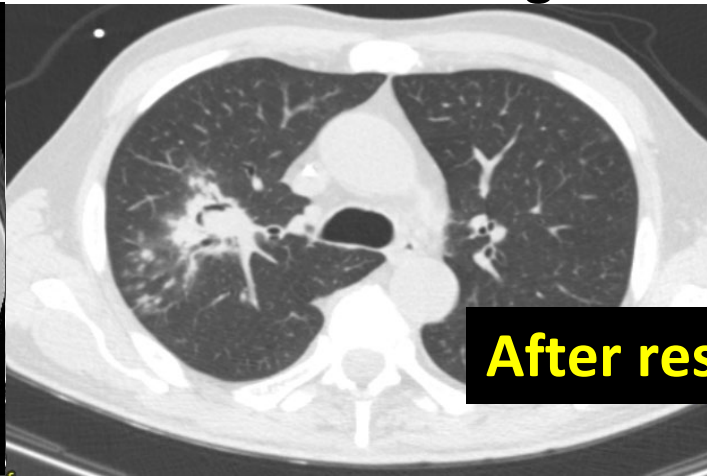


# EORCT/MSG 2019 Clinical criteria for pulmonary IMI

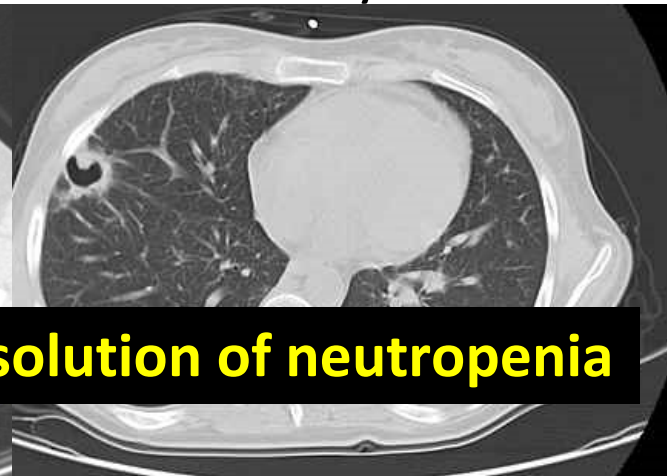
Nodule +/-halo sign



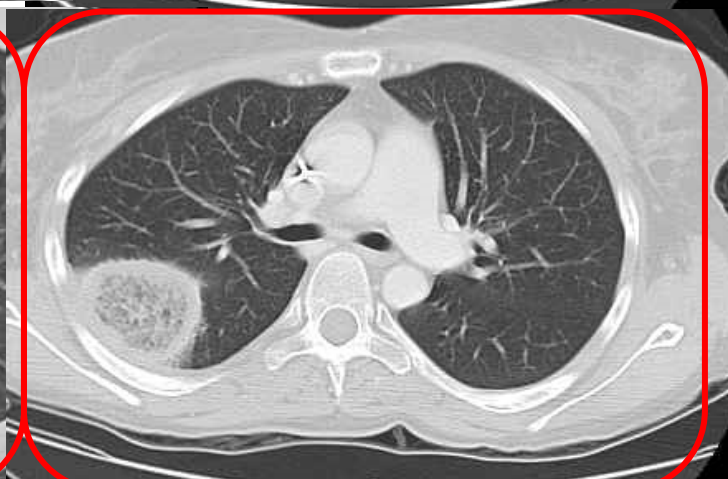
Air crescent sign



Cavity



**After resolution of neutropenia**



Wedge-shape and segmental or lobar **consolidations**

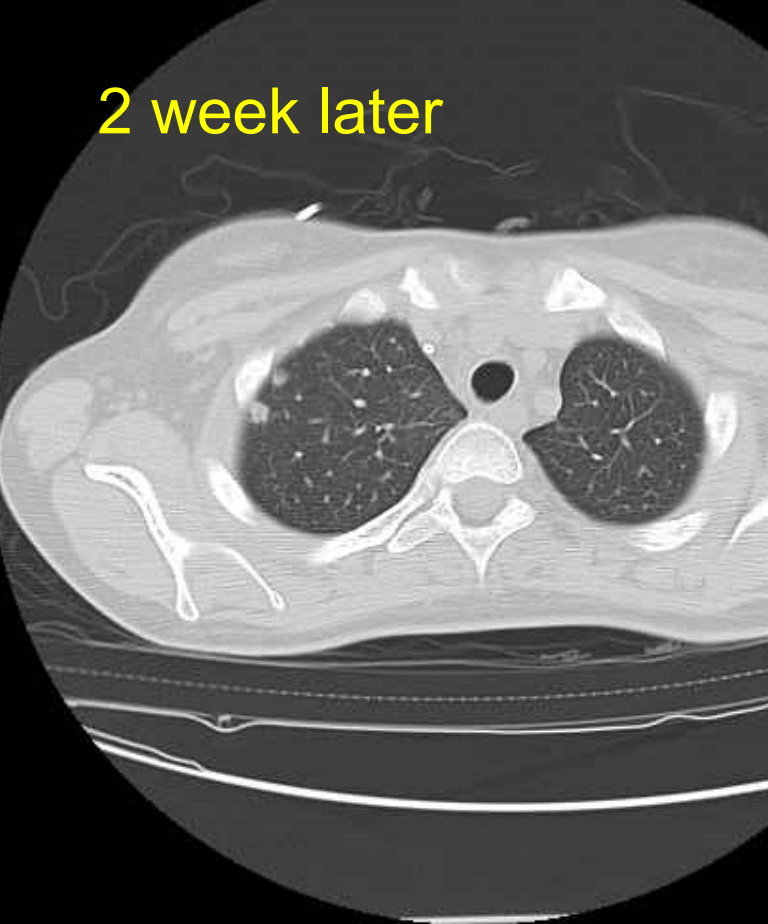
**Reverse halo sign**



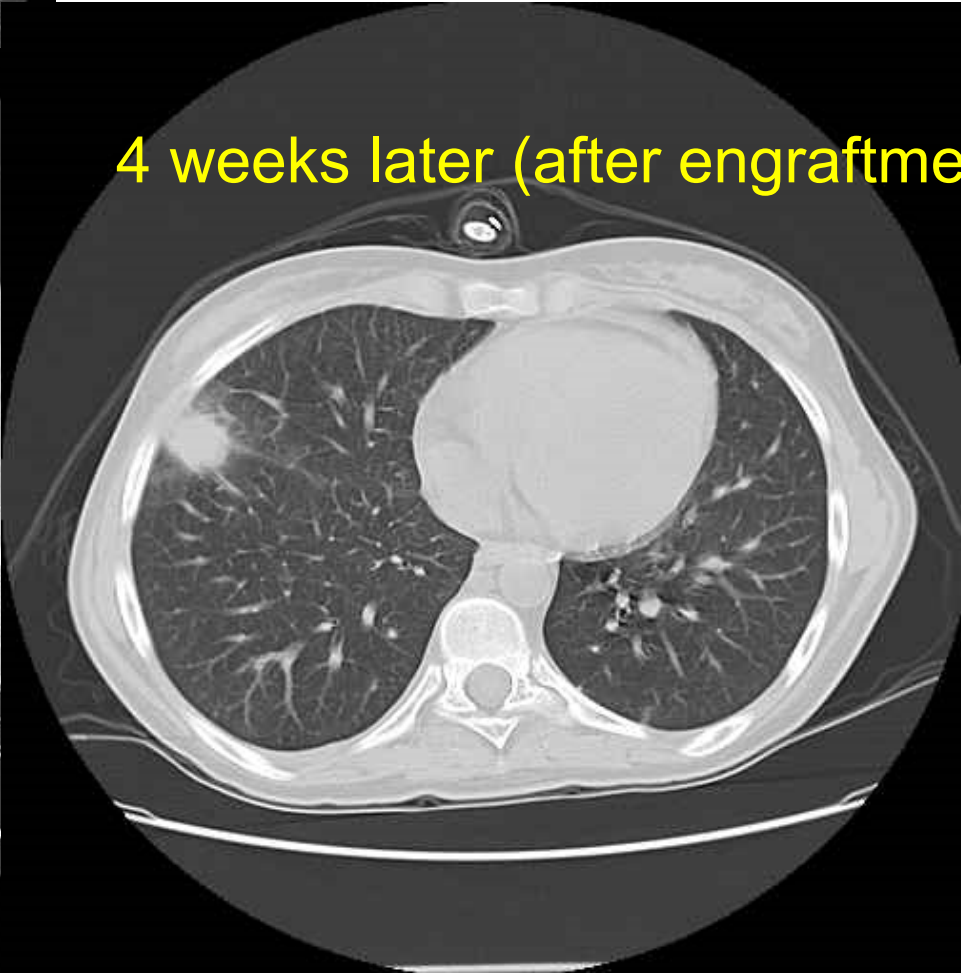
## with the wisdom of hindsight

19 yo female with SAA undergoing second alloHSCT  
Long term neutropenia, no mould active prophylaxis  
GM screening positive 0.6

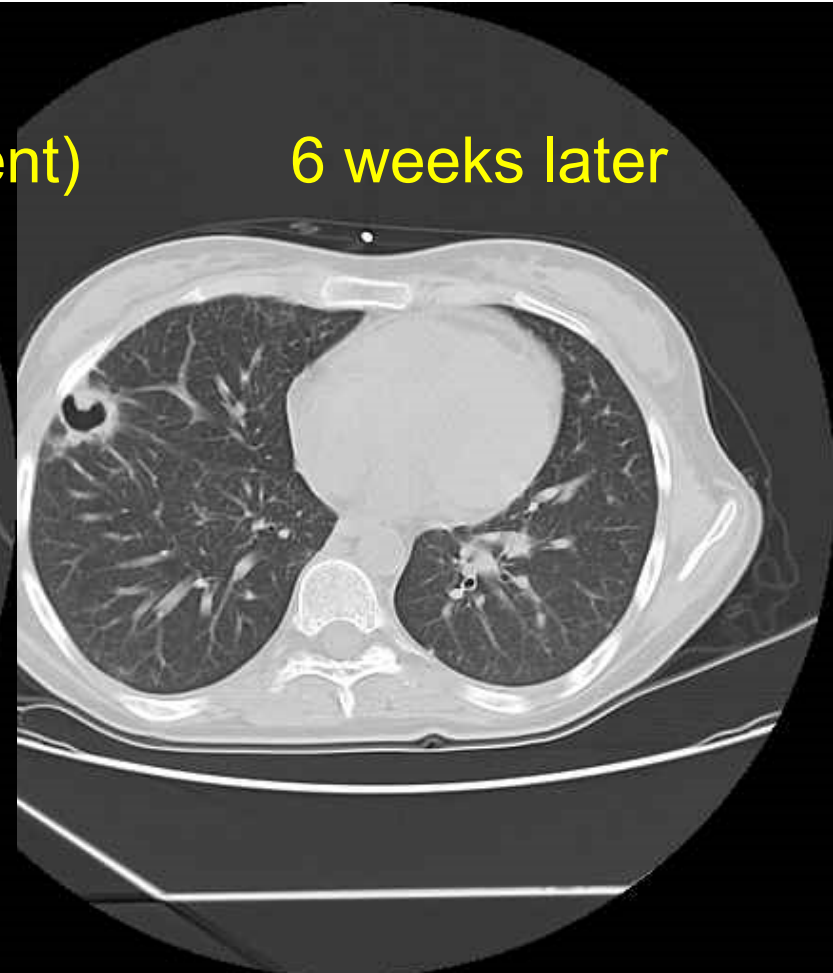
2 week later



4 weeks later (after engraftment)



6 weeks later



# EORTC/MSGERC criteria were designed for clinical trials and epidemiology

Not for everyday clinical decisions

**blood** 2012 119: 1831-1837  
 Prepublished online October 18, 2011;  
 doi:10.1182/blood-2011-04-351601

The strategy for the diagnosis of invasive pulmonary aspergillosis should depend on both the underlying condition and the leukocyte count of patients with hematologic malignancies

Anne Bergeron, Raphaël Porcher, Annie Sulahian, Cédric de Bazelaire, Karine Chagnon, Emmanuel Raffoux, Anne Vekhoff, Muriel Cornet, Françoise Isnard, Benoît Brethon, Claire Lacroix, Jean Louis Poirot, Claire Bouges, Francis Derouin, Abdelatif Tazi and Patricia Ribaud

**Table 1.** Patterns of invasive fungal disease in practice, based on 2008 EORTC-MSG criteria.

	A	B	C				D	E
	-	-	I	II	III	IV	-	
<b>Radiological signs and clinical symptoms</b>	No	Persistent febrile neutropenia	No	Clinical (any new infiltrate not fulfilling the EORTC/MSG criteria)		Radiological signs on CT (dense, well-circumscribed lesion(s) with or without a halo sign, air-crescent sign, or cavity)		Not considered necessary
<b>Mycology results</b>	Negative	Negative	Positive biomarker or microscopy or culture	Negative	Positive biomarker or microscopy or culture	Negative	Positive biomarker or microscopy or culture	Positive tissue or specimen from a sterile site
<b>Clinical evidence of IFD</b>	No	No	No	No	No	Yes	Yes	Yes
<b>Mycological evidence of IFI</b>	No	No	Yes	No	Yes	No	Yes	Yes
<b>Final diagnosis</b>	Unclassified					Possible IMD	Probable IMD	Proven IMD



**Figure 1.** Lung CT scans from 2 different patients. (A) Nodule with a halo sign revealing angioinvasive aspergillosis in an AL patient. (B) Centrilobular nodules and tree-in-bud opacities showing airway-invasive aspergillosis in a patient who underwent an allogeneic HSCT.

# Performance of GM in BAL, the role of cut-off

Cut-off value	No. of participants	Summary estimates (95% CI)	Implications	Quality and Comments
0.5 ODI	229 with (12 studies)	Sensitivity: 0.88 (95% CI 0.75 to 1.00)	At a prevalence of 12%, 120 out of 1000 patients will develop proven or probable IA. Of these, 14 will be missed.	All studies except one had a high risk of bias in two or more domains.
1.0 ODI	177 with (11 studies)	Sensitivity: 0.78 (95% CI 0.61 to 0.95)	At a prevalence of 12%, 120 out of 1000 patients will develop proven or probable IA. Of these, 26 will be missed.	For all studies the concerns regarding applicability were low.
			894 with possible or no IA	Of the 880 patients with possible or no IA, 167 will have a false-positive test result and may be unnecessarily treated. Of all 273 patients with a positive test, 106 will indeed have proven or probable IA; of negative tests, 14 will have IA after all (2%).
			594 with possible or no IA	Of the 880 patients with possible or no IA, 6 test result and may be unnecessarily treated. Of all 156 patients with a positive test, 94 will indeed have proven or probable IA; of all 844 negative tests, 26 will have IA after all (3%).
interpretation and extrapolation of these results has to be			Note, the populations and results were very heterogeneous. Therefore, performed with caution.	

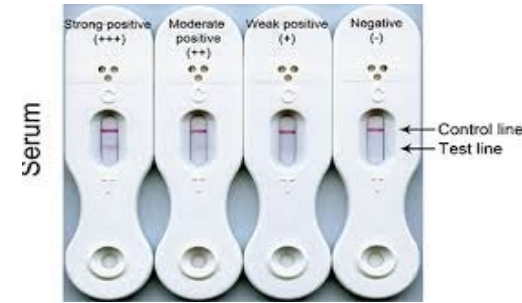
# Faster GM results - Towards point-of-care testing

- **Reliable, quantitative «almost» PoC for BAL fluid (with pre-treatment): 15–25 minutes, two assays available:**<sup>1,2</sup>

1. Aspergillus galactomannan **LFA**
2. Aspergillus-specific **LFD** that detects the mannoprotein antigen secreted by growing Aspergillus with JF5 MAbs

- A 2015 meta-analysis investigated LFD in BAL:<sup>2</sup>
  - Sensitivity: 86% (95% CI: 76–93)
  - Specificity: 93% (95% CI: 89–96)
- Included in the 2018 ESCMID IA guidelines (B II)<sup>3</sup>

- Qualitative/semi-quantitative results were provided initially > optical reader
- Compared with visual readouts, digital readouts provide quantitative results and perform better with BAL (235 samples from HM patients) for both assays – LFA and LFD<sup>1</sup>
- There was a good correlation with traditional GM for LFA<sup>5</sup>



# *Aspergillus* PCR: blood and BAL

- Included in some 2018 ESCMID, 2019 EORTC/MSG guidelines
- **In addition to in-house methods, commercial assays are available**
  - For *A. fumigatus* or for several species, not all differentiate between species
  - Some detects also **mutations associated with azole-resistance in *A. fumigatus*** (TR<sub>34</sub>/L98H, T289A, Y121F)
- Initial studies: sensitivity 68-94%, specificity 80-98%
- **Recent reports: sensitivity not specificity seems a crucial issue**
  - Sensitivity/specificity 30%/91%; 40%/69%; 65%/100%
- The performance variable, higher in culture positive samples
  - Our experience: sensitivity for proven/probable IA in H
    - 40% with one commercially available assay
    - 92% with another

# Clinical impact of PCR-based *Aspergillus* and azole resistance detection in invasive aspergillosis. A prospective multicenter study.

## Unexpected lessons

- Prospective study in the Netherlands and Belgium
- Evaluated the clinical value of the multiplex AsperGenius®PCR in hematology patients from 12 centers
- This PCR detects the most frequent *cyp51A* mutations in *A. fumigatus* conferring azole-resistance
- **Inclusion: patients with a CT-scan showing a pulmonary infiltrate and BAL within 48h (=possible IA)**
- The primary endpoint - antifungal treatment failure in patients with azole-resistant IA
- 323 patients enrolled (32% alloHSCT), complete mycological and radiological information available in 94% (276/323)
- **Probable IA – 36% (99/276)**
- PCR testing in BAL in 91% (293/323): **positive in 40% (116/293), with *A. fumigatus* DNA in 30% (89/293)**
- The resistance PCR was conclusive in 65% (58/89) and resistance detected in 8/58 (14%)

**Table 2. Microbiology Results Including Bronchoalveolar Fluid Galactomannan, AsperGenius Polymerase Chain Reaction, and Culture**

	BALf GM		
	<0.5	0.5–0.99	≥1
Number of patients (n) <sup>a</sup>	215	32	74
Aspergenius performed	193	31	68
PCR <i>Aspergillus</i> species–positive	50 (26%)	16 (52%)	50 (74%)
PCR <i>Aspergillus</i> species–negative	143 (74%)	15 (48%)	18 (26%)
PCR <i>Aspergillus fumigatus</i> –positive	38 (20%)	12 (39%)	39 (57%)
PCR <i>Aspergillus fumigatus</i> –negative	156 (80%)	19 (61%)	29 (43%)
PCR <i>Aspergillus terreus</i> –positive	1 (0.5%)	0 (0%)	2 (3%)

# Clinical impact of PCR-based Aspergillus and azole resistance detection in invasive aspergillosis. A prospective multicenter study.

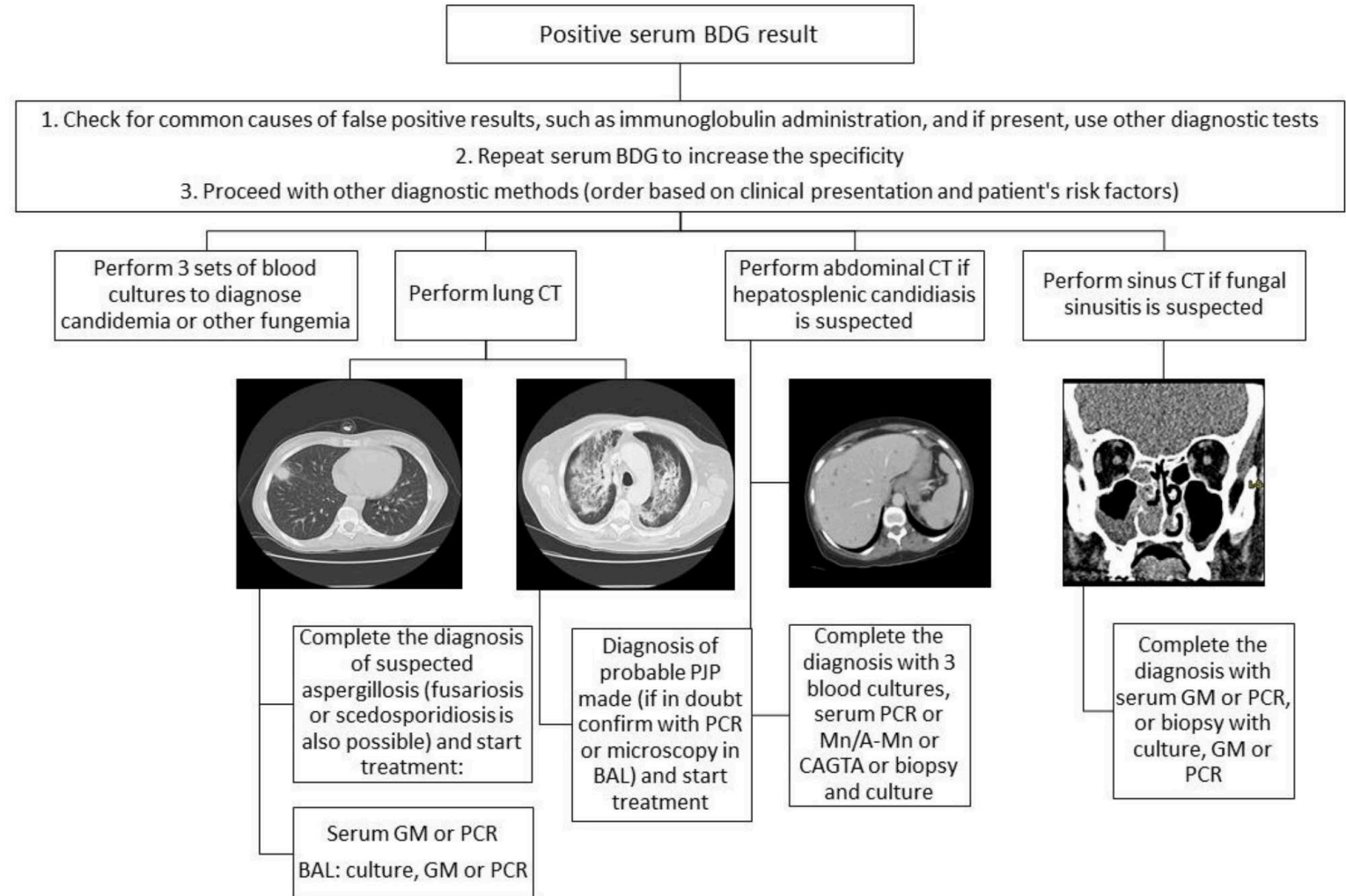
	GM pos (N=77)	Culture pos (N=24)	PCR pos (N=119)	PCR pos in duplicate (N=67)	GM and culture neg but PCR pos (N=62)	<b>GM and culture neg but PCR pos in duplicate (N=28)</b>	GM, culture and PCR neg (N=154)
Antifungal therapy started at BAL (-5, +14 days) (n/N)	72/77 (94%)	23/24 (96%)	105/119 (88%)	62/67 (93%)	52/62 (84%)	24/28 (86%)	105/154 (68%)
Median (IQR) days of antifungals	27 (11 – 73)	38 (17 – 88)	32 (10 – 89)	33 (12 – 89)	34 (10 – 123)	71 (15 – 135)	18 (7 – 63)
<b>6-week mortality (n/N)</b>	<b>23/76 (30%)</b>	<b>8/24 (33%)</b>	<b>26/119 (22%)</b>	<b>16/67 (24%)</b>	<b>9/62 (15%)</b>	<b>4/28 (14%)</b>	<b>24/153 (16%)</b>

BAL galactomannan positivity was associated with higher mortality (p=0.004)

**Mortality of patients with an isolated positive Aspergillus PCR in BAL was comparable to those with a negative PCR (p=0.83)**



# BDG – to be used in combination only



# Specifics of IFD in haematology

## Prophylaxis

High rate (>6-8%) of IMI in certain patient groups but not in others, thus anti-mould prophylaxis strongly recommended in certain groups  
Breakthrough IFD present, also due to persistent immune deficit

## Diagnosis

EORTC/MSG diagnostic criteria available for 20 years and updated, but not developed for daily clinical practice (e.g. bronchoinvasive aspergillosis – unclassified)

## Prognosis

High mortality, particularly if ongoing severe immunosuppression > fear of rapid deterioration + **suboptimal diagnosis > leading to empirical therapy**



# Empirical antifungal therapy

## 40 years ago

Definition: antifungal treatment in **neutropenic** patients with persistent fever despite 4-7 days of broad-spectrum antibiotics, or in patients with relapsing fever

Reason: IFD during neutropenia is impossible to diagnose:

low yield of cultures,

late radiological signs on X-ray

and is associated with high mortality

Pizzo et al. Am J Med 1982 (16 vs 18 patients)

EORTC Am J Med 1989 (64 vs 68 patients)

## Today



Widely available **galactomannan**, with short turn-around time (use of **point-of-care tests** in selected cases)



**Rapidly available CT** (same day)



Available **Aspergillus-PCR** in serum and BAL, serum glucan (IC, PjP) with short turn-around time



Rapidly available **BAL** – to avoid false negative results due to antifungal treatment (Aspergillus PCR might be more helpful than GM)



Rapidly available cerebral RM



> Only as time-buying strategy  
Diagnostic availability is critical

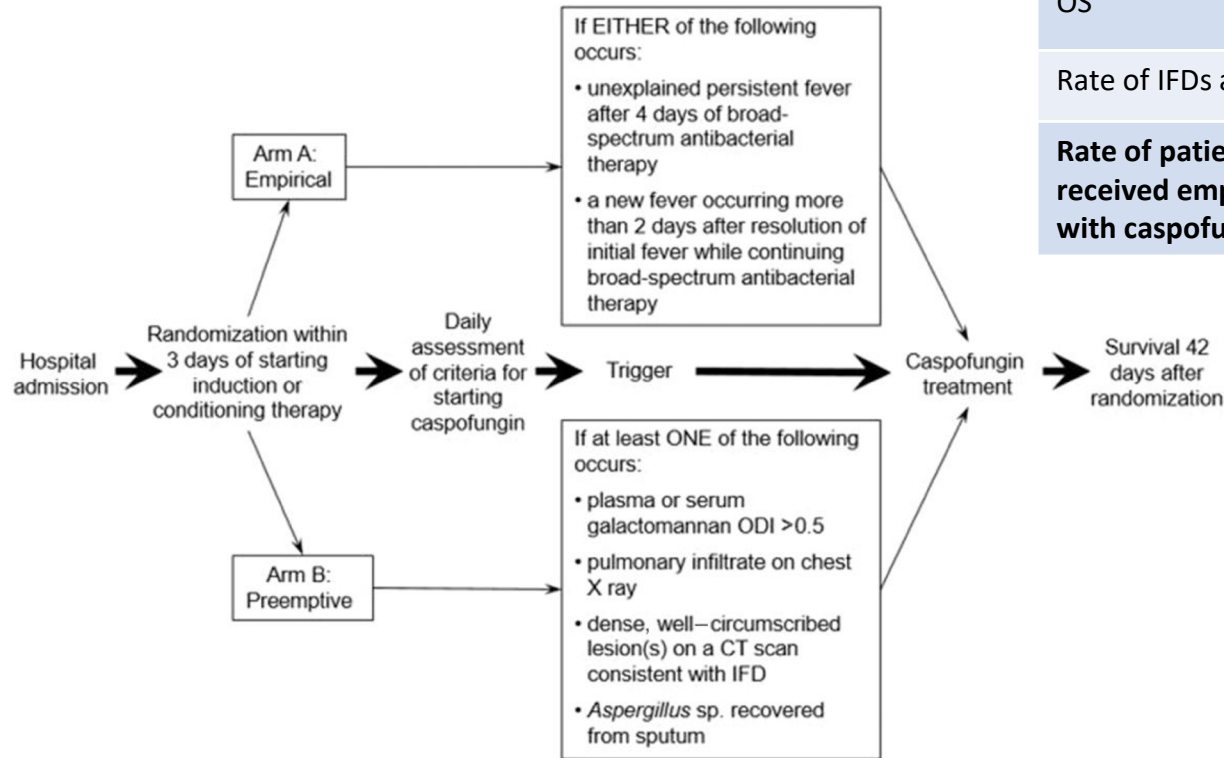


# Empiric vs Preemptive Antifungal Strategy in High-Risk Neutropenic Patients on Fluconazole Prophylaxis: A Randomized Trial of the European Organization for Research and Treatment of Cancer

Johan Maertens,<sup>1</sup> Tom Lodewyck,<sup>2</sup> J. Peter Donnelly,<sup>3</sup> Sylvain Chantepie,<sup>4</sup> Christine Robin,<sup>5</sup> Nicole Blijlevens,<sup>3</sup> Pascal Turlure,<sup>6</sup> Dominik Selleslag,<sup>2</sup> Frédéric Baron,<sup>7</sup> Mickael Aoun,<sup>8</sup> Werner J. Heinz,<sup>9</sup> Hartmut Bertz,<sup>10</sup> Zdeněk Ráčil,<sup>11</sup> Bernard Vandercam,<sup>12</sup> Lubos Drgona,<sup>13</sup> Valerie Coiteux,<sup>14</sup> Cristina Castilla Llorente,<sup>15</sup> Cornelia Schaefer-Prokop,<sup>3</sup> Marianne Paesmans,<sup>8</sup> Lieveke Ameye,<sup>8</sup> Liv Meert,<sup>16</sup> Kin Jip Cheung,<sup>16</sup> Deborah A. Hepler,<sup>17</sup> Jürgen Loeffler,<sup>18</sup> Rosemary Barnes,<sup>19</sup> Oscar Marchetti,<sup>20,21</sup> Paul Verweij,<sup>3,6</sup> Frederic Lamoth,<sup>20</sup> Pierre-Yves Bochud,<sup>20</sup> Michael Schwarzwinger,<sup>22</sup> and Catherine Cordonnier<sup>3</sup>, for the Infectious Diseases Group and the Acute Leukemia Group of the European Organization for Research and Treatment of Cancer

2012-2015

pre-emptive: twice weekly galactomannan screening and CT scan on demand  
549 included, 80% AML, 20% alloHSCT



	Pre-emptive, % (95%CI)	Empirical, % (95%CI)	P
OS	96.7% (93.8%–98.3%)	93.1% (89.3%–95.5%)	NS
Rate of IFDs at day 84	7.7% (4.5%–10.8%)	6.6% (3.6%–9.5%)	NS
<b>Rate of patients who received empirical treatment with caspofungin</b>	<b>27%</b>	<b>63%</b>	<b>&lt;.001</b>

Figure 1. Study design. Abbreviations: CT, computed tomography; IFD, invasive fungal disease; ODI, optical density index

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## Prognosis

High mortality, particularly if ongoing severe immunosuppression > fear of rapid deterioration + **suboptimal diagnosis** > **leading to empirical therapy**

## Prophylaxis

High rate (>6-8%) of IMI in certain patient groups but not in others, thus anti-mould prophylaxis strongly recommended in certain groups  
Breakthrough IFD present, also due to persistent immune deficit

## Treatment

Drug-drug interactions between triazoles and numerous anti-neoplastic drugs – need for TDM  
Frequent hepatic toxicity due to numerous causes (chemotherapy, hepatic Graft vs. Host Disease)  
Optimal length of IMD treatment ? (until the resolution of immune deficit...)

# Invasive aspergillosis. First line treatment

	ECIL-6 2016	IDSA 2016 (Strength of recommendation and evidence)	ESCMID – ECMM – ERS 2018	Australia 2021
Voriconazole	A I (oral CIII) TDM indicated	Strong. High quality	A I	A I
Isavuconazole	A I	Alternative to voriconazole A II	A I	A I
Posaconazole	-	-	-	A I*
L-AMB 3mg/kg	B I	Strong. Moderate quality	B II	B II
ABLC 5mg/kg	B II	Weak. Low quality	C III	-
ABCD	C I	Weak. Low quality	D I	-
D-AMB	A I against use	-	D I	-
Caspofungin	C II	Not recommended	C II	C II
Micafungin	-	Weak. Moderate quality	C III	C II
Anidulafungin	-	-	-	-
Itraconazole	C III	-	C III	-
Voriconazole + anidulafungin	C I	Weak. Moderate quality	C I	C I
Other combinations	C III	-	D III	-

Antifungal agents

Fosmanogepix	Ibrexafungerp	Olorofim	Opelconazole	Rezafungin
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Pathogens

 <i>Aspergillus calidoustus</i>	Green	Green	Green	Green
<i>Aspergillus fumigatus</i>	Green	Green	Green	Green
Azole-resistant <i>A. fumigatus</i>	Green	Green	Red	Green
<i>Aspergillus flavus</i>	Green	Green	Green	Green
<i>Aspergillus lentulus</i>	Green	Green	Green	Green
<i>Aspergillus nidulans</i>	Green	Green	Green	Green
<i>Aspergillus niger</i>	Green	Green	Red	Green
<i>Aspergillus terreus</i>	Green	Green	Green	Green
<i>Aspergillus tubingensis</i>	Green	Green	Green	Green

 <i>Cunninghamella</i>	Orange	Red	Red	Green
<i>Lichtheimia</i>	Orange	Red	Red	Green
<i>Mucor</i>	Orange	Red	Red	Green
<i>Rhizopus</i>	Orange	Red	Red	Green

 <i>Fusarium spp.</i>	Green	Red	Orange
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 <i>Alternaria alternata</i>	Orange	Green	Red
<i>Cladosporium spp.</i>	Green	Green	Green
<i>Paecilomyces variotii</i>	Green	Orange	Green
<i>Purpureocillium lilacinum</i>	Green	Red	Orange
<i>Scopulariopsis spp.</i>	Green	Red	Green
<i>Rasamsonia spp.</i>	Green	Green	Green

 <i>Scedosporium spp.</i>	Green	Orange	Green
<i>Lomentospora prolificans</i>	Green	Orange	Green




# When Primary Antifungal Therapy Fails

**Table 1. Causes of antifungal therapy failure.**

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Causes of antifungal therapy failure

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Host factor	
Severity of illness	
Persistence of immunodeficiency (e.g., neutropenia or use of corticosteroids)	
Primary (intrinsic) drug resistance	
Wrong diagnosis	
Mixed infection	
Low concentration of the drug at the site of infection	
Pharmacokinetic and pharmacodynamic	
Drug interactions	
Biofilms	
Poor vascular supply (e.g., abscess and necrotic tissue)	
Drug toxicities (direct and with drug interactions)	
Development of resistance (secondary)	
Misdiagnosis of failure—immune reconstitution inflammatory syndrome	

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