



Approccio Diagnostico-terapeutico e Criticità nella Gestione delle VAP

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Francesco G. De Rosa

Associate Professor, Infectious Diseases

University of Turin, Italy

Fellow, Infectious Diseases Society of America

Disclosures

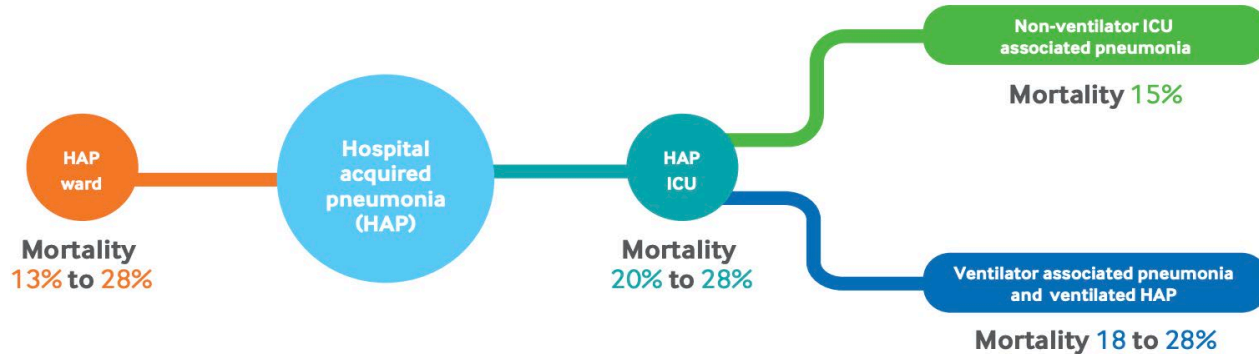
Consultant/advisory board/speaker fees

- Pfizer, MSD, Angelini, Tillots, Menarini
- Thermo Fisher, Shionogi
- BioTest, Nordic Pharma, InfectoPharma
- Gilead Sciences, GSK, Hikma, Advanz, Correvio

Research grant

- Pfizer, MSD, Shionogi

Hospital-acquired Pneumonia



- **Incidence: 5–40%**
- Progressive risk: peak at **5–9 days of MV**
- **Metrics depending on setting and diagnostic algorithm**
- Microbiological confirmation: strongly recommended; method: controversial
- **Prevention:** reduction of modifiable & non-modifiable risk factors
- **Treatment duration:** 7 days, with caveat
- **De-escalation & antibiotic-free** days to be implemented

Definitions

- **CDC/NHSH 2008¹**
- **CDC 2012**
- **ECDC-HELICS 2010²** Clinical + imaging + microbiology
- **CDC/NHSH 2013-2021:³**
 - *VAE: ventilator-associated events;*
 - *VAC: ventilator-associated condition;*
 - *IVAC: infection-related ventilator-associated complication;*
 - *Possible and probable VAP*

Ventilator-associated events (VAE):

This definition is for use in surveillance; it is not a clinical definition algorithm and is not intended for use in the clinical management of patients

1. Horan TC *et al.* *Am J Infect Control* 2008;36:309–32; 2. Plachouras D *et al.* *Intensive Care Med* 2018;44:2216–18; 3. Centres for Disease Control and Prevention (2018) National Healthcare Safety Network (NHSN) Patient Safety Component Manual. CDC, Atlanta. Available at: https://www.cdc.gov/nhsn/pdfs/pscmanual/pscmanual_current.pdf (accessed September 2023).

HAP & VAP

- “Disease Difficult to Characterize” -

No Diagnostic gold standard: Diagnostic Methodology

Invasive Vs. Non-invasive

Conventional Vs. fast-track

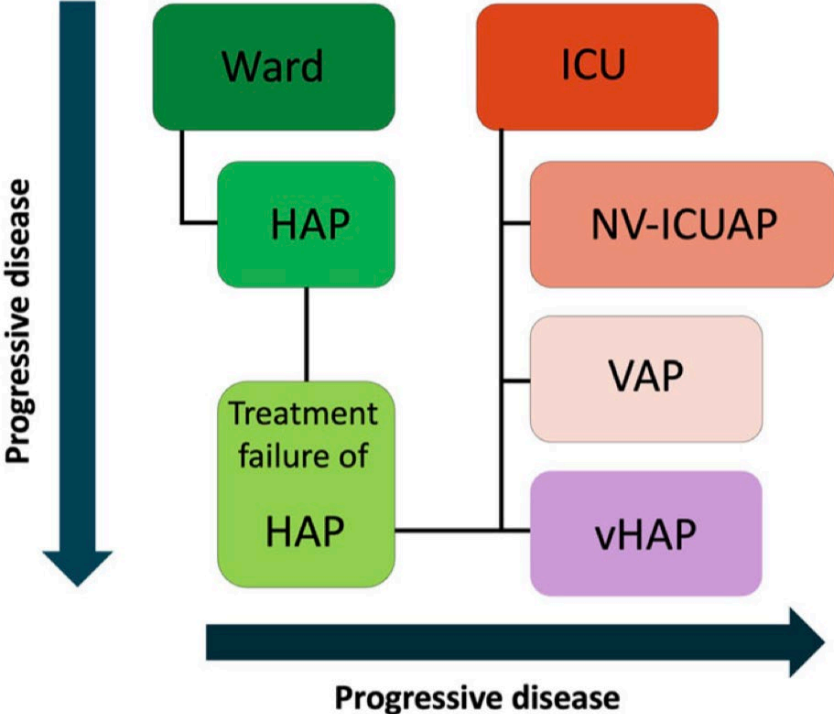
Algorithms of empiric treatment

MRSA, Enterobacterales (ESBL, KPC, NDM)

P. aeruginosa, A. baumannii

Example of "Progressive Disease" in HAP - VAP

Amani Alnimir Infect Dis Ther 2023



Risk Factors for HAP & VAP

European Vs. US Guidelines

Table 2 | High risk of MDR pathogens in HAP/VAP

European guidelines (2017)	US guidelines (2016)
Previous antibiotic treatment	Previous antibiotic treatment
≥5 days of hospitalization	≥5 days of hospitalization
Septic shock	Septic shock
Hospital settings with high rates of MDR pathogens (>25%)	ARDS before VAP
Previous colonization by MDR Pathogens	Acute renal replacement therapy before initiation of VAP
Mortality risk >15%	

VAP=ventilator associated pneumonia; ARDS=acute respiratory distress syndrome; HAP=hospital-acquired pneumonia; MDR=multidrug resistant.^{4,6}

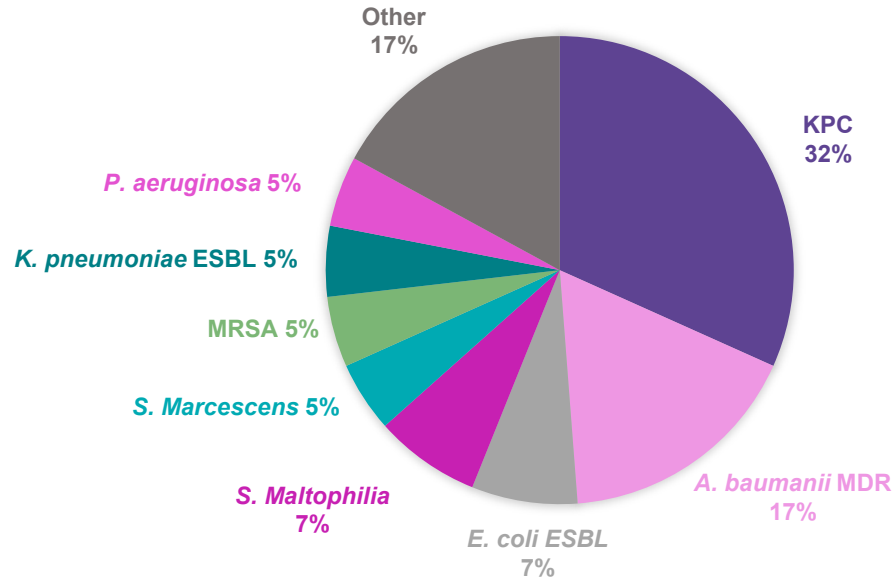
Risk Factors for HAP & VAP

MDRO	Risk factors
MRSA	<ul style="list-style-type: none">⇒ Age⇒ NP appearance > 6 days after admittance⇒ NP development excluding summers⇒ Respiratory diseases⇒ Multilobar involvement
<i>Pseudomonas aeruginosa</i>	<ul style="list-style-type: none">⇒ Admission to ICU, antimicrobial use⇒ Prior carbapenem⇒ Invasive operation⇒ Previous non-KPC-Kp infections⇒ Duration of previous antibiotic therapy before KPC colonization
<i>Enterobacteriaceae</i>	<ul style="list-style-type: none">⇒ Male sex⇒ Admission from another health care facility⇒ Ventilation at any point before culture during the index hospitalization⇒ Receipt of any carbapenem in the prior 30 days⇒ Receipt of any anti-MRSA agent in the prior 30 days
<i>Acinetobacter baumannii</i>	<ul style="list-style-type: none">⇒ APACHE II score at admission⇒ Systemic illnesses (chronic respiratory disease and cerebrovascular accident)⇒ Presence of excess non-invasive or invasive devices (mechanical ventilation)⇒ Ever used antibiotics within 28 days (carbapenem and cefepime)

Etiology of HAP-VAT-VAP (2010–2019)

Reference	Type of infection	Microbiology				
Ferrer et al. [30]	HAP	<i>S. aureus</i> , 17.7%	<i>P. aeruginosa</i> , 17.7%	<i>E. coli</i> , 6.5%	<i>Enterobacter</i> spp., 4.3%	<i>K. pneumoniae</i> , 3.2%
Esperatti et al. [22]	VAP	<i>P. aeruginosa</i> , 24%	<i>S. aureus</i> , 23%	<i>E. coli</i> , 7%	<i>Enterobacter</i> spp., 6%	<i>H. influenzae</i> , 4%
Restrepo et al. [31]	VAP	<i>S. aureus</i> , 38.7%	<i>H. influenzae</i> , 23.4%	<i>P. aeruginosa</i> , 14.7%	<i>k. pneumoniae</i> , 11.5%	<i>E. coli</i> , 11.1%
		MDR, 30%				
Quartin et al. [32]*	VAP	<i>S. aureus</i> , 60.3%	<i>P. aeruginosa</i> , 9.4%	<i>Acinetobacter</i> spp., 7.3%	<i>Klebsiella</i> spp., 6.8%	<i>Enterobacter</i> spp., 5.1%
Nseir et al. [33]	VAT	<i>P. aeruginosa</i> , 34.4%	<i>S. aureus</i> , 20.5%	<i>A. baumannii</i> , 11.5%	<i>K. oxytoca</i> , 10.6%	<i>Enterobacter</i> spp., 9.8%
		MDR, 36.8%				
Martín-Loeches et al. [21]	VAT	<i>P. aeruginosa</i> , 25%	<i>S. aureus</i> , 23%	<i>Klebsiella</i> spp., 15%	<i>E. coli</i> , 12%	<i>Enterobacter</i> spp., 11%
		MDR, 61%				
	VAP	<i>P. aeruginosa</i> , 24%	<i>S. aureus</i> , 24%	<i>Klebsiella</i> spp., 14%	<i>Enterobacter</i> spp., 12%	<i>E. coli</i> , 11%
		MDR, 61%				
ECDC [18]	VAP	<i>P. aeruginosa</i> , 20.8%	<i>S. aureus</i> , 17.8%	<i>Klebsiella</i> spp., 16.1%	<i>E. coli</i> , 13.3%	<i>Enterobacter</i> spp., 10.3%
Koulenti et al. [29]	HAP	<i>Enterobacteriaceae</i> , 32.9%	<i>S. aureus</i> , 24.9%	<i>P. aeruginosa</i> , 17.4%	<i>A. baumannii</i> , 15.4%	
ENVIN-HELICS [3]	VAP	<i>P. aeruginosa</i> , 23.8%	<i>S. aureus</i> , 13.5%	<i>Klebsiella</i> spp., 10.3%	<i>E. coli</i> , 9.1%	<i>Enterobacter</i> spp., 8.6%
		PIP/TAZ R, 34.1%	MRSA, 12.7%	PIP/TAZ R, 50%	PIP/TAZ R, 21.7%	
		Carba R, 37.9%		Carba R, 23.5%	Carba R, 0%	
		Collistin R, 8.6%		3 ^o G cef R, 3.7%	3 ^o G cef R, 12.5%	
Pulido et al. [34]	VAP	<i>P. aeruginosa</i> , 21.1%	<i>A. baumannii</i> , 17.9%	<i>K. pneumoniae</i> , 15.6%	<i>S. aureus</i> , 13.3%	<i>E. coli</i> , 7.8%
Huang et al. [35]	VAP	<i>A. baumannii</i> , 33.9%	<i>K. pneumoniae</i> , 23.6%	<i>P. aeruginosa</i> , 19.8%	<i>S. aureus</i> , 7.1%	<i>S. maltophilia</i> , 3.8%
		Carba R, 76.4%	Carba R, 44%	Carba R, 59.5%	MRSA, 60%	
Cantón-Bulnes et al. [36]	VAT	<i>P. aeruginosa</i> , 24.5%	<i>H. influenzae</i> , 18.9%	<i>E. coli</i> , 9.4%	<i>S. aureus</i> , 9.4%	<i>K. pneumoniae</i> , 7.5%
Ibn Saied et al. [37]	VAP	<i>P. aeruginosa</i> , 33.5%	<i>Enterobacteriaceae</i> , 32.3%	<i>S. aureus</i> , 19%	<i>S. pneumoniae</i> , 4.9%	<i>S. maltophilia</i> , 4.7%

VAP: Example of Local Epidemiology on Respiratory Isolates



Further changes expected with NDM Enterobacterales

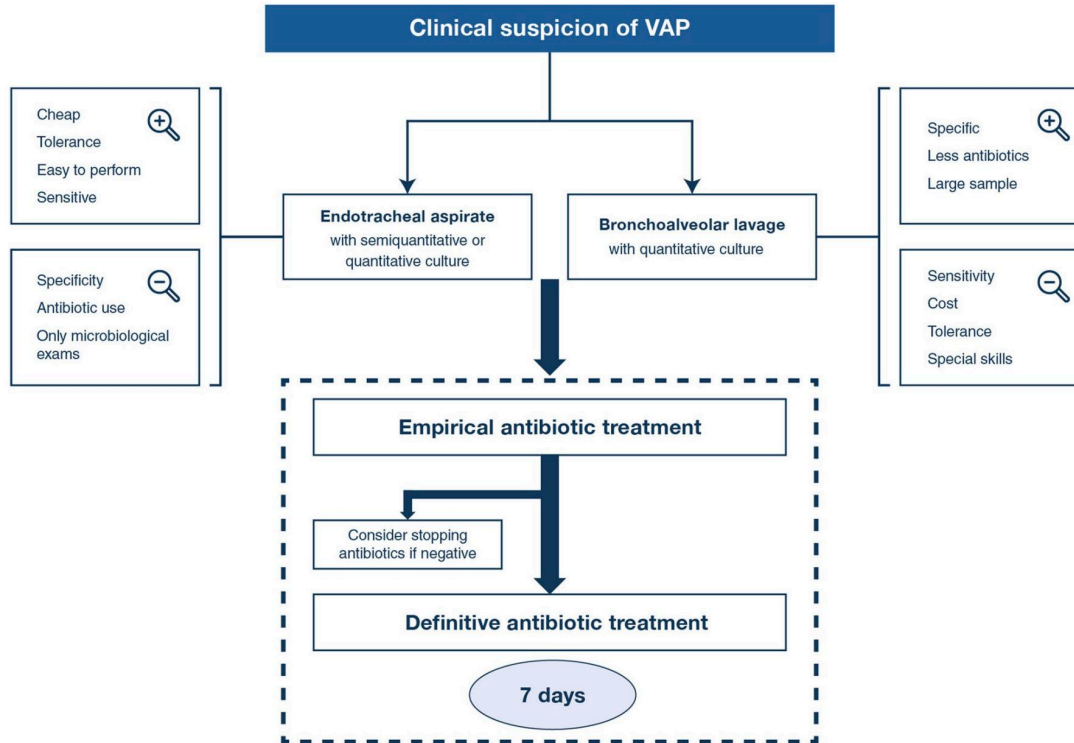
Diagnosis of VAP in Critically Ill Adult Patients: A Systematic Review & Meta-analysis

Estimates of the performance of physical examination, chest radiography, laboratory values, and CPIS for the diagnosis of ventilator-associated pneumonia, relative to **reference standard of histopathology from lung biopsy**

	No. of cohorts (patients)	Sensitivity, % (95% CI)	Specificity, % (95% CI)	Diagnostic odds ratio (95% CI)	Positive likelihood ratio (95% CI)	Negative likelihood ratio (95% CI)	GRADE
<i>Histopathology reference standard</i>							
Fever	5 (142)	66.4 (40.7–85)	53.9 (34.5–72.2)	2.31 (0.98–5.43)	1.44 (1.01–2.05)	0.62 (0.36–1.09)	Low
Purulent secretions	4 (336)	77 (64.7–85.9)	39 (25.8–54)	2.13 (1.34–3.41)	1.26 (1.06–1.5)	0.59 (0.42–0.83)	Moderate
Infiltrate on chest radiography	7 (238)	88.9 (73.9–95.8)	26.1 (15.1–41.4)	2.83 (1.18–6.82)	1.2 (1.03–1.4)	0.42 (0.2–0.92)	Low
Leukocytosis	3 (88)	64.2 (46.9–78.4)	59.2 (45–72)	2.6 (1.05–6.45)	1.57 (1.03 to 2.4)	0.61 (0.36–1.01)	Low
Sputum from endotracheal aspirate (> 10 ⁵ CFU/mL)	3 (75)	75.7 (51.5–90.1)	67.9 (40.5–86.8)	6.59 (2.17–20.04)	2.36 (1.19–4.66)	0.36 (0.18–0.73)	Very Low
Protected specimen brush (> 10 ³ CFU/mL)	7 (201)	61.4 (43.7–76.5)	76.5 (64.2–85.6)	5.19 (2.31–11.65)	2.62 (1.63–4.19)	0.5 (0.33–0.77)	Low
Bronchoalveolar lavage (> 10 ⁴ CFU/mL)	10 (307)	71.1 (49.9–85.9)	79.6 (66.2–88.6)	9.57 (4.04–22.71)	3.48 (2.13–5.7)	0.36 (0.2–0.66)	Low
CPIS > 6	4 (343)	73.8 (50.6–88.5)	66.4 (43.9–83.3)	5.56 (1.30–23.84)	2.2 (1.09–4.43)	0.4 (0.17–0.92)	Low

CFU = colony-forming units; CI = confidence interval; CPIS = Clinical Pulmonary Infection Score; GRADE = Grading of Recommendations, Assessments, Development and Evaluation

VAP: Clinical & Microbiological Strategies



“Invasive Microbiology”

2016 Clinical Practice Guidelines by the Infectious Diseases Society of America and the American Thoracic Society [14]

2017 International ERS/ESICM/ESCMID/ALAT guidelines [18]

Should patients with suspected VAP be treated based on the results of invasive sampling (BAL, PSB, blind mini-BAL) with quantitative culture results, noninvasive sampling (endotracheal aspiration) with quantitative culture results, or noninvasive sampling with semiquantitative culture results?

Recommendation

We suggest noninvasive sampling with semiquantitative cultures to diagnose VAP, rather than invasive sampling with quantitative cultures and rather than noninvasive sampling with quantitative cultures (weak recommendation, low-quality evidence)

If invasive quantitative cultures are performed, should patients with suspected VAP whose culture results are below the diagnostic threshold for VAP (PSB with $< 10^3$ CFU/mL, BAL with $< 10^4$ CFU/mL) have their antibiotics withheld rather than continued?

Recommendation

Noninvasive sampling with semiquantitative cultures is the preferred methodology to diagnose VAP; however, the panel recognizes that invasive quantitative cultures will occasionally be performed by some clinicians. For patients with suspected VAP whose invasive quantitative culture results are below the diagnostic threshold for VAP, we suggest that antibiotics be withheld rather than continued (weak recommendation, very low-quality evidence)

In intubated patients suspected of having VAP, should distal quantitative samples be obtained instead of proximal quantitative samples?

Recommendation

We suggest obtaining distal quantitative samples (prior to any antibiotic treatment) in order to reduce antibiotic exposure in stable patients with suspected VAP and to improve the accuracy of the results. (weak recommendation, low quality of evidence)

We recommend obtaining a lower respiratory tract sample (distal quantitative or proximal quantitative or qualitative culture) to focus and narrow the initial empiric antibiotic therapy. (strong recommendation, low quality of evidence)

BAL Bronchoalveolar lavage, PSB protected specimen brush, CFU colony-forming units

French Multicenter Study: Clinical Vs. Br-BAL/PSB

Fagon & Chastre, Ann Intern Med 2000; 132: 621

Multicentric RCT of VAP outcomes; 413 pts in 31 ICUs

Clinical Dx (n=204) Vs. Br-PSB/BAL (n=209)

Br-BAL/PSB used to adjust or stop Rx

Microbiological Methodology & Analysis

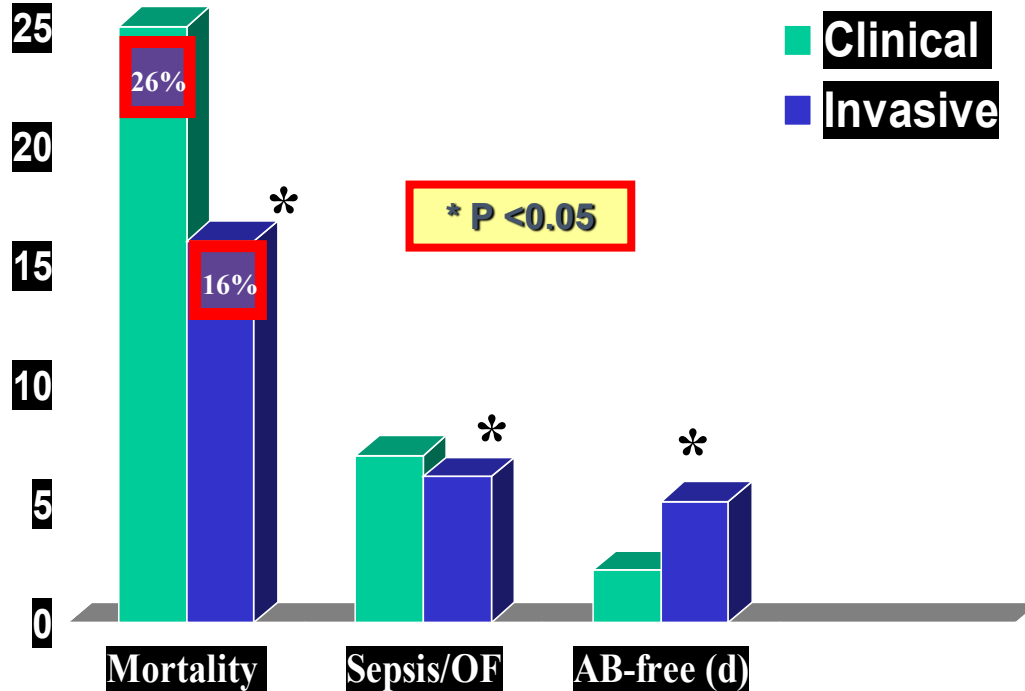
Fagon & Chastre, Ann Intern Med 2000; 132: 621

- **Clinical group: Endotracheal Aspirate**
 - Sterile collection by a suction catheter, vortexed for 1 minute
 - Gram staining and culture

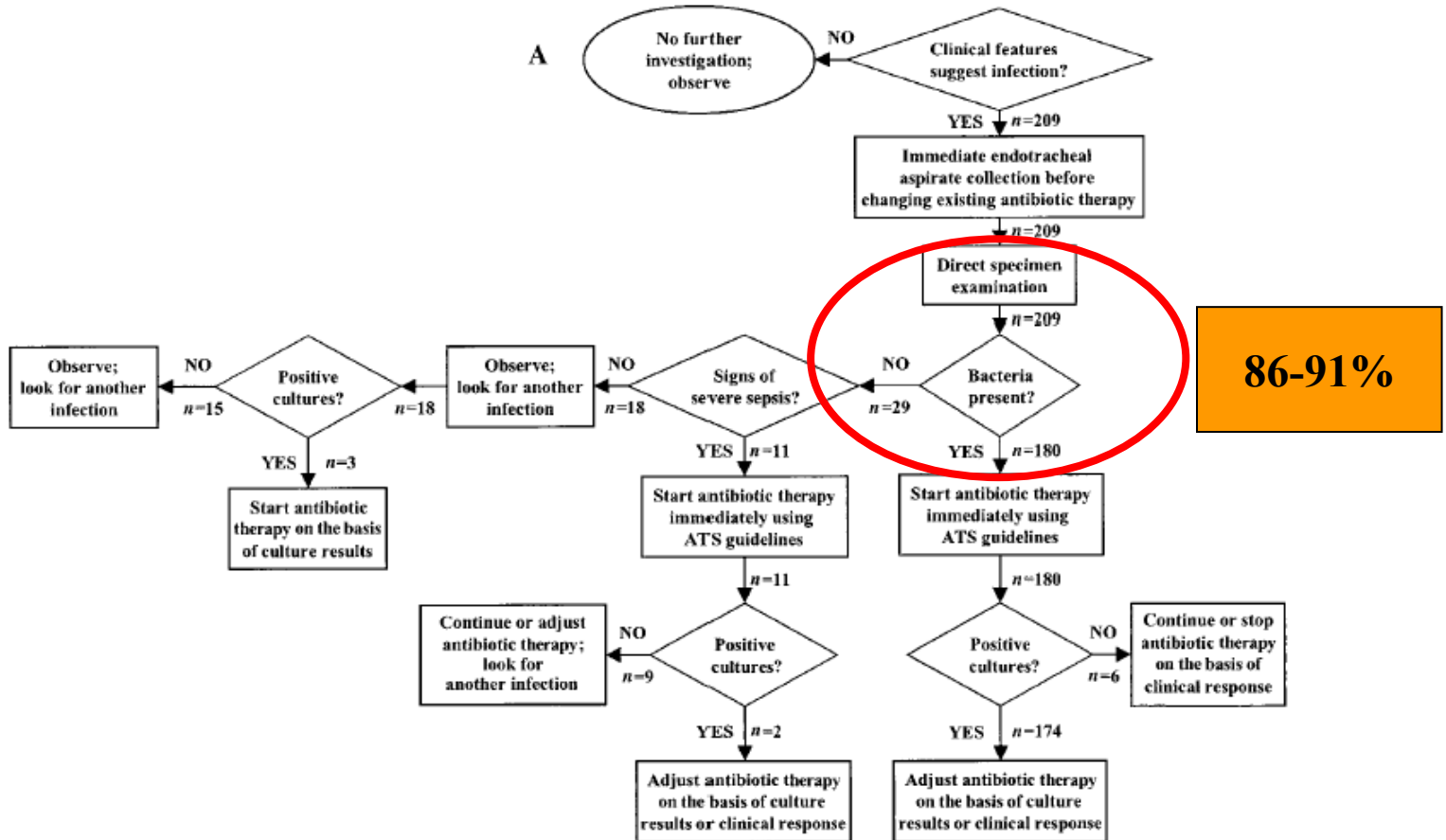
- **Invasive group**
 - **PSB or BAL**
 - **One sample:**
 - Gram staining of cytocentrifuge preparations
 - % of cells containing intracellular bacteria
 - **One sample:**
 - Quantitative cultures

Outcomes

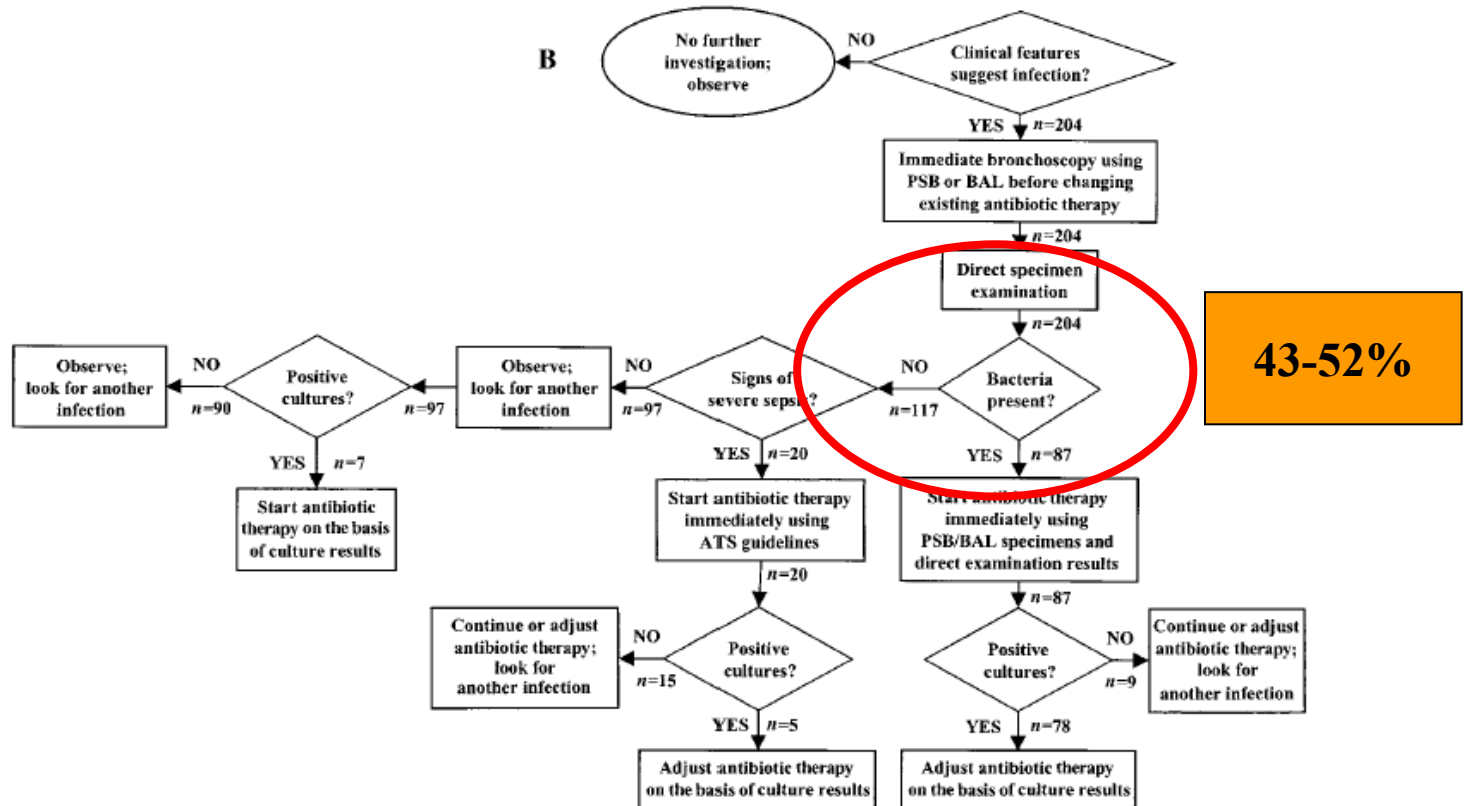
Fagon & Chastre, Ann Intern Med 2000; 132: 621



Clinical Group

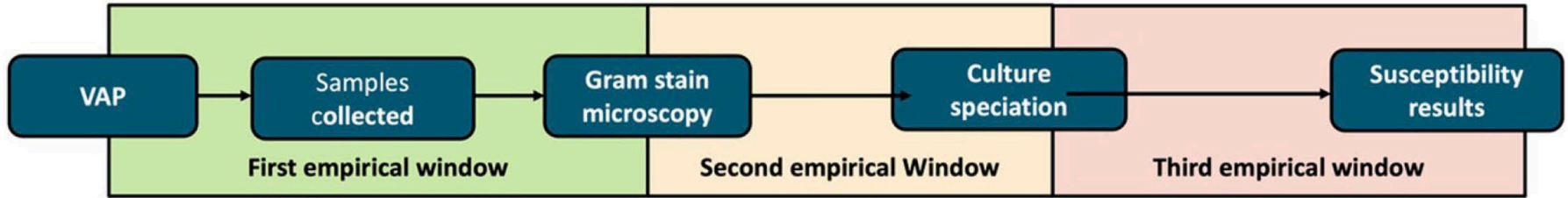


Invasive Group



Empiric Therapeutic Window in VAP

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Nosocomial Pneumonia in the Era of Multidrug-Resistance: Updates in Diagnosis and Management

Elena Xu ¹, David Pérez-Torres ^{2,†} , Paraskevi C. Fragkou ^{3,†}, Jean-Ralph Zahar ^{4,†} and Despoina Koulenti ^{1,5,*} 

Table 2. Advantages and disadvantages of multiplex PCR panels.

Advantages	Disadvantages
Exceptionally faster time to results for pathogen and resistance profiles: major utility for prompt treatment modification and effective patient management	Over-detection of microbial and viral genome: problem in results interpretation: pathogen or coloniser? (may be partially solved with semi-quantification of bacterial targets)
Multiple targets detection at the same and Detection of viral and atypical pathogens as well	The presence of a resistance gene marker may not be linked to the detected microorganism, but to other co-existent organisms either undetectable or below the detection limit, thus making culture-based techniques still necessary in many cases
Detection of pathogens even when antimicrobial treatment has been initiated	Initial cost to buy the equipment
Potential for better antibiotic utilisation and positive impact on: -nosocomial pneumonia management, shortening hospital stay and decreasing healthcare costs, -antibiotic stewardship programs	Not widely available among different institutions yet
Early identification of MDR pathogens should facilitate enhanced infection control practices and reduce spread	Further validation versus traditional diagnostic techniques needed and determination of the effect on antimicrobial prescribing, patient outcomes and resistance is needed

Quantitative Agreement: Bacterial Targets Between PN Panel And SOC

TABLE 3 Quantitative agreement of bacterial targets

PN panel result (copies/ml)	No. of samples ^a with SOC culture result (CFU/ml)			
	Not detected	10 ³	10 ⁴	≥10 ⁵
Not detected	3,734	3	0	0
10 ⁴	24 (8)	6	4	0
10 ⁵	27 (17)	3	4	1
10 ⁶	9 (4)	2	12	3
≥10 ⁷	13 (7)	2	15	23
% concordant^b	98.1 (3,734/3,807)	18.8 (3/16)	11.4 (4/35)	100 (27/27)

^aNumbers in parentheses are the numbers of culture-negative results obtained for specimens from patients who received antibiotics with potential activity against the given bacterial target detected within 72 h preceding specimen collection. One laboratory reported bacterial culture quantitation (11 isolates) as “few,” “moderate,” or “many”; these were categorized as 10³, 10⁴, and ≥10⁵ CFU/ml, respectively. Shading indicates concordance between the BioFire PN panel and routine culture quantitation.

^bConcordance between the PN panel and culture quantitation among all positive cultures was 43.6% (34/78).

In positive cultures, concordance = 43.6% (34/78)

- Concordance poorest with low bacteria
- **Strong bias toward higher semiquantitative values**, when PN panel is compared to culture
- In case of **discordant quantification**, the **PN panel result was higher than that of culture**, with 72.2% (34/44) of the results **exceeding culture quantification by >1 log**

- **Targets quantified as 10⁴ copies/ml by the PN panel may frequently be quantified as 10³ CFU/ml by routine culture**, ie the culture-based limit of detection
- Their clinical significance is largely unreported
- Interpretation should be done in the context of other clinical & laboratory results (e.g., additional pathogens present at high concentrations in the same specimen)
- **All bacterial targets reported as 10⁵ CFU/ml in culture were reported as 10⁵ genomic copies/ml by the PN panel**

Early Use of PN Panel → Early Adaptation of Therapy & Reduction of Unnecessary Antimicrobial Exposure & in Adult Patients with Pneumonia

Retrospective multicenter study in 4 French University Hospitals.

159 adult patients with a suspicion of CAP, HAP and **VAP**, with appropriate samples (mainly ETA) available. 30 pts not ICU located

Assess the impact of PN+ on early adaptation of empirical AB therapy in adult pneumonia patients;

A group of experts (ICU, ID, Lab) examines clinical data and PN+ result to recommend an AB treatment without knowing the culture results for these patients. → **simulates the real-life situation of having PN+ results BEFORE culture**

Table 4 Impact of the rm-PCR results on antibiotic prescription, according to the multidisciplinary committee ($n = 159$)

	Overall, $n = 159$	CAP, $n = 54$	HAP, $n = 68$	VAP, $n = 37$
Antibiotic modification	123 (77)	37 (69)	54 (79)	32 (87)
De-escalation	63 (40)	20 (37)	25 (37)	18 (49)
Escalation	35 (22)	8 (15)	18 (27)	9 (24)
Undetermined	25 (16)	9 (17)	11 (16)	5 (14)
No change	36 (23)	17 (32)	14 (21)	5 (14)

Results are presented as n (%)

CAP community-acquired pneumonia, HAP hospital-acquired pneumonia, VAP ventilator-associated pneumonia

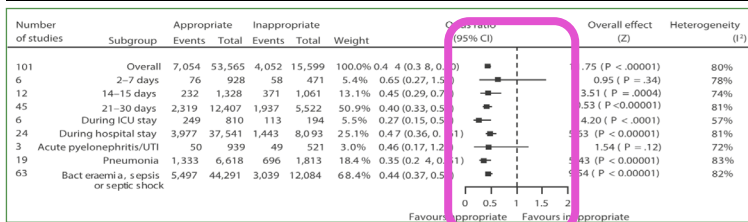
ATB treatment would have been changed in 77% of the cases on average (VAP: 87%);

De-escalation of empiric AB therapy would be the predominant treatment change;

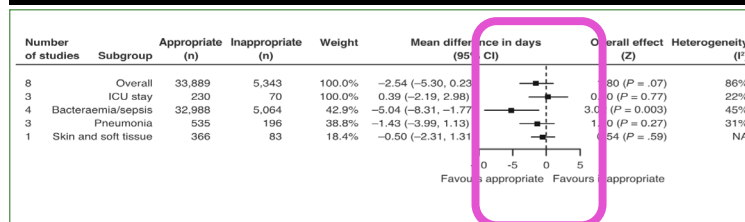
Adoption of PN+ would have decreased use of β -lactams from 92% to 82% ('main treatment') and β -lactam companion therapies from 50% to 31%.

Appropriate Vs. Inappropriate Antimicrobial Therapy: Mortality, Cost-effectiveness & Health-economic Outcomes

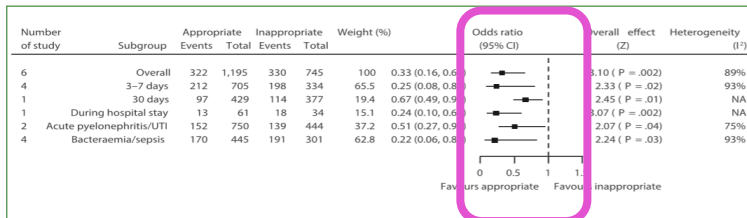
Mortality



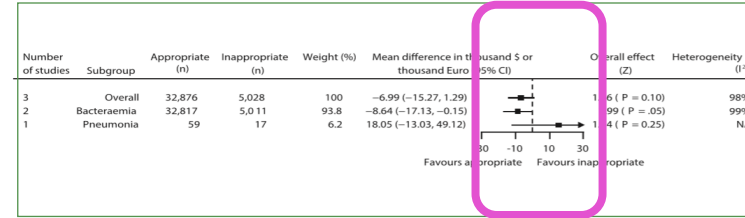
Length of Hospital Stay



Treatment Failure



Costs



Rapid diagnostics for early identification of etiology ensure appropriate initial antimicrobial therapy

VAP: Initial Antibiotic Treatment

Situation	Therapeutic class	Agent
Early VAP (< 5 days), without MDR bacteria risk factor*	Non-antipseudomonal β -lactam	Amoxicillin/clavulanic acid OR Third generation cephalosporin
Late VAP (\geq 5 days), OR Risk factors for MDR bacteria	β -lactam active against <i>Pseudomonas aeruginosa</i> AND Non β -lactam antipseudomonal agent	Cefepime 2 g q 8 h OR Ceftazidime 2 g q 8 h OR Piperacillin–tazobactam 4 g q 6 h OR Meropenem 2 g q 8 h Amikacin 25 mg/kg/day OR Ciprofloxacin 1200 mg/day
Known MRSA colonization, or high (> 20%) MRSA prevalence in the unit	Agent active against MRSA	Vancomycin 30–45 mg/kg/day OR Linezolid 600 mg/12 h
Known colonization with carbapenem-resistant <i>Enterobacteriaceae</i> or <i>Pseudomonas aeruginosa</i> susceptible only to new beta-lactam agents	New β -lactam agent	Ceftolozane–tazobactam 3 g q 8 h [†] OR Ceftazidime–avibactam 2.5 g q 8 h [†] OR Meropenem–vaborbactam 4 g q 8 h [†] OR Imipenem–relebactam 1.5 g q 6 h [†]

MDR Risk Factors

- Antibiotics: previous 90 days
- Hospital stay >5 days
- Septic shock at VAP onset
- ARDS prior to VAP onset
- RRT prior to VAP onset
- Previous MDR colonization

*This situation and the corresponding antimicrobial agents are not mentioned in IDSA/ATS guidelines

[†]The empirical use should be restricted to patients colonized by specific pathogens (carbapenem-resistant *Enterobacteriaceae* or extensively drug-resistant *P. aeruginosa*), according to previous susceptibility testing

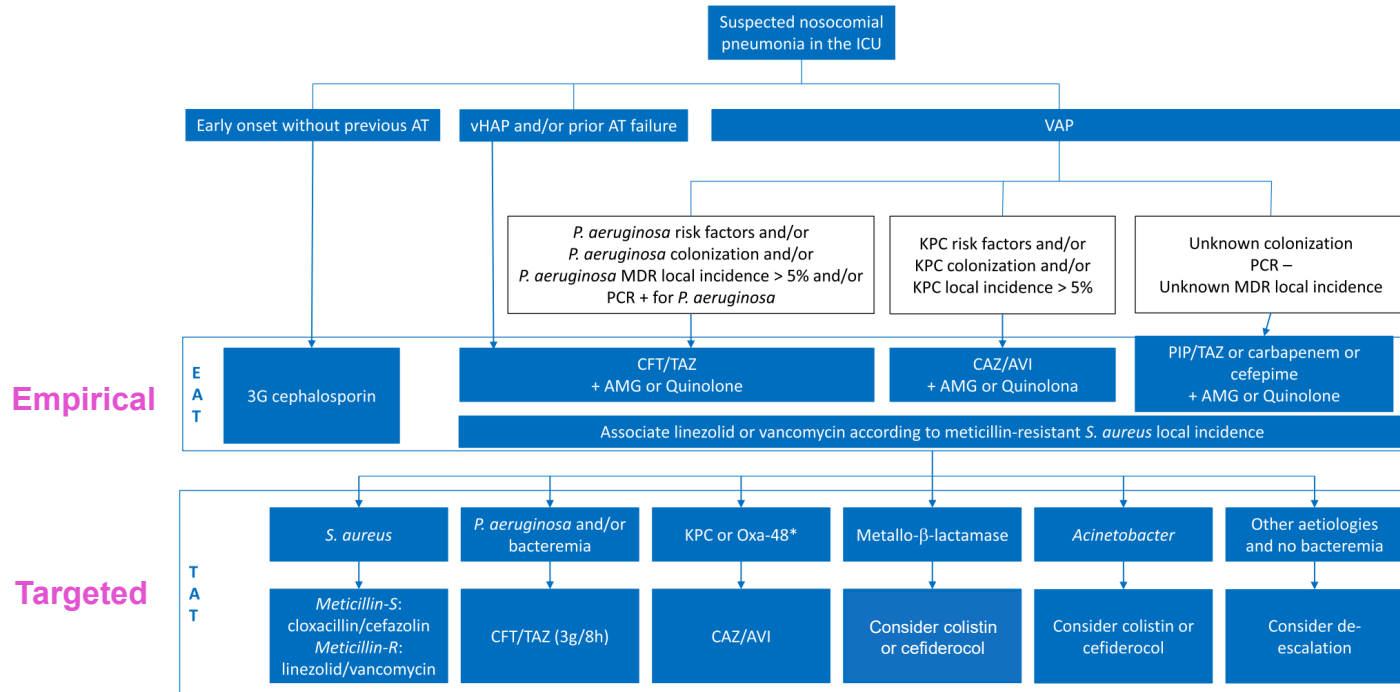
US & European Guidelines: Choice of Empiric Treatment

Table 1 | Empiric treatment according to US And European guidelines

Risk for MDR pathogens	US guidelines	European guidelines
Low		(≤15% mortality risk, low MDR risk) Narrow spectrum antimicrobial with activity for meticillin susceptible <i>S aureus</i> and non-resistant Gram negatives: ertapenem, ceftriaxone, cefotaxime, moxifloxacin, levofloxacin
High		(>15% mortality risk and/or high MDR risk) No septic shock: monotherapy with broad spectrum agent active against >90% of likely Gram negative pathogens +/- MRSA (if > 25% of <i>S aureus</i> isolates are MRSA) Septic shock: combination therapy with anti-pseudomonal regimen +/- MRSA (if > 25% of <i>S aureus</i> isolates are MRSA)

Both guidelines agree on stratifying patients according to individual and local community risk factors for MDR pathogens.⁶

Treatment Algorithm: Example



Wide Window for the new BL/BLI, including Cefiderocol

vHAP, ventilated hospital-acquired pneumonia

Zaragoza et al. Crit Care 2020;24:383.

Duration of Antibiotic Therapy in Gram-negative Infections with a Particular Focus on MDR pathogens

Ongoing studies on shorter <8 days treatment of VAP

REGARD-VAP trial

- Multinational multicentre study, 460 patients enrolled (November 22, 2023)
- Aims: clinical non-inferiority and superiority of SAT (up to 7 days) vs long antibiotic treatment (LAT)
- NCT03382548: <https://clinicaltrials.gov/ct2/show/NCT03382548>

DATE trial (Duration of Antibiotic Treatment for Early VAP)

- Completed in early 2022 with pending results: 22 patients enrolled
- 4- vs 8-day treatment for early VAP
- NCT01994980: <https://clinicaltrials.gov/ct2/show/NCT01994980>

Discontinuation of Therapy: IDSA Guidelines

Recommendation:

1. For patients with HAP/VAP, who have been on antimicrobial therapy for 7 days or more, use the following criteria to guide the discontinuation of antibiotic therapy (weak recommendation, low-quality evidence)

**De-escalation
Highly recommended
- IDSA & ERS Guidelines -**

Remarks:

It is not known if the benefits of using PCT levels to determine whether or not to discontinue antibiotic therapy exist in settings where standard antimicrobial therapy for VAP is already 7 days or less

Respiratory Viruses in Nosocomial Pneumonia: An Evolving Paradigm

Nosocomial spread of common respiratory viruses

Common cause of nosocomial pneumonia

- Influenza, respiratory syncytial virus, adenovirus, and rhinovirus
- Especially among immunosuppressed and pediatric patients

Difficult differential between bacterial & viral infections

Crude mortality in viral HAP and VAP

Rival or exceed those in bacterial NP

Current medical needs

- Rigorous prospective, multicenter trials
- Studies of novel therapeutics for these viral infections

Metagenomics for Microbiological Diagnosis of HAP & VAP

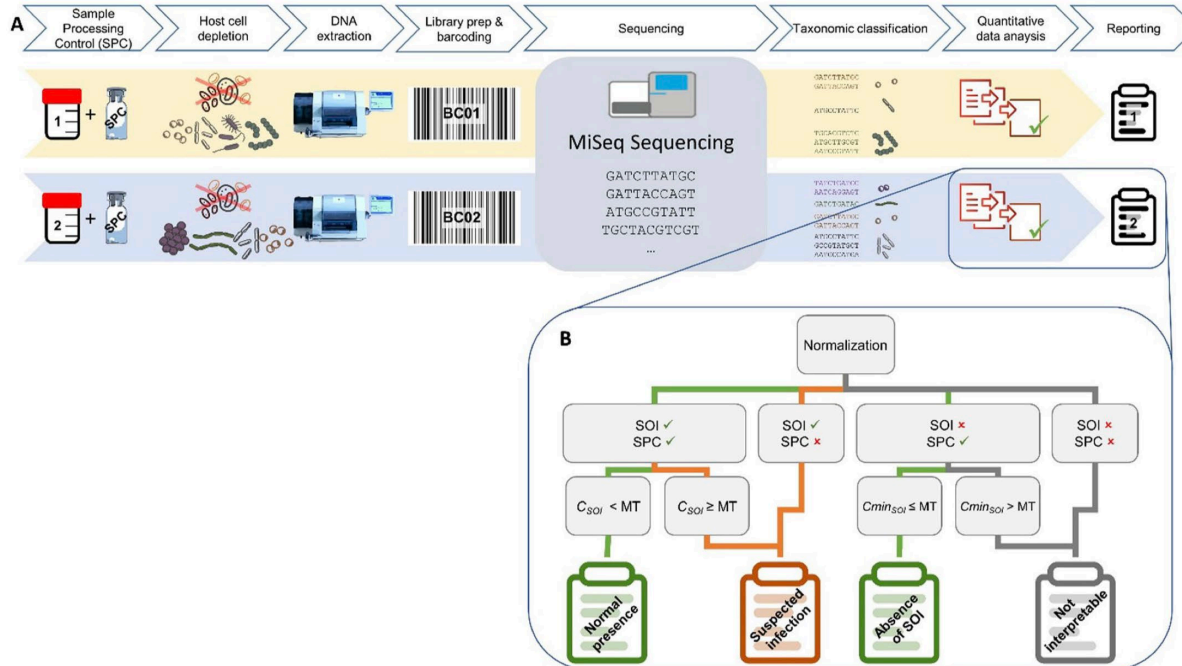


Fig. 1 Complete workflow for clinical metagenomic analysis of BAL samples. **A** is the experimental workflow in which two independent samples are analyzed in the same sequencing run. **B** represents the rule of interpretation applied independently to each SOI to determine whether it is involved in patient infection or presence at normal concentration or absence in the sample or the inability to interpret the result. *SOI* species of interest, *SPC* sample processing control, *MT* metagenomic threshold

Metagenomics for Microbiological Diagnosis of HAP & VAP

Heitz M et al. Resp Research 2023

Table 1 Microorganisms of the mNGS pneumonia panel (n = 19)

Acinetobacter baumannii

Citrobacter freundii

Citrobacter koseri

Enterobacter aerogenes

Escherichia coli

Haemophilus influenzae

Hafnia alvei

Klebsiella oxytoca

Klebsiella pneumoniae

Legionella pneumophila

Morganella morganii

Proteus mirabilis

Proteus vulgaris

Providencia stuartii

Pseudomonas aeruginosa

Serratia marcescens

Staphylococcus aureus

Stenotrophomonas maltophilia

Streptococcus pneumoniae

(Early) Sensitivity Vs. (Delayed) Specificity

A. Esame microscopico diretto / (Fast track) / colturale

B. Decidere su quali casi usare Fast-track Microbiology

- Criteri diagnostico-terapeutici
- Protocollo condiviso con microbiologi, intensivisti e pneumologi

C. Fast-track

- Escalation (myne)– de-escalation

D. Definite de-escalation "composita" con colturali (CIII?)

E. Integrare uso biomarkers

F. Antibiotic free-days importante misura in ICU

Conclusions

- **Choice of clinical / invasive strategy**
- **Fast microbiological testing for VAP**
 - Potential results in real time to tailor therapy on pathogens and MDR determinants
 - Support clinical data
- **Local ICU ecology of utmost importance**
- **Mathematical models and scoring systems**
 - Still not sufficiently developed for operational application
- **Areas for priority research studies in VAP**
 - Clinical utility & implementation of rapid diagnostics
 - Validation of prediction scores in making clinical decisions